

Fusobacterium is toxic for head and neck squamous cell carcinoma and its presence may determine a better prognosis

Head and neck squamous cell carcinoma (HNSCC) is a devastating disease. Despite morbid treatment, 5-year survival rates remain poor (28%-67%) [1]. There is a significant knowledge gap regarding how the microbiota may impact HNSCC treatment efficacy [2]. We used microbiome data from two independent cohorts to test and validate the hypothesis that oral bacteria are associated with HNSCC prognosis and in vitro models to investigate mechanistic underpinnings. Methods are detailed in Supplementary Materials.

We first explored associations between the relative abundance (RA) of bacterial genera and overall survival (OS) time in 155 patients with mucosal HNSCC available in the Cancer Microbiome Atlas (TCMA, Supplementary Table S1, Supplementary Text). The distribution of bacterial genera is shown in Supplementary Figure S1. Linear stepwise and Cox regression modeling evaluated associations between these genera and OS/DSS. Only Fusobacterium detectability was associated with both better OS (hazard ratio [HR] = 0.35, 95% confidence interval [CI] = 0.15-0.83], P = 0.018, Supplementary Figure S2A) and better disease-specific survival (DSS; 0.28 [0.15-0.83], P =0.031, Supplementary Figure S2B). Kaplan-Meier survival analysis mirrored these results (Figure 1A-B). Additionally, Fusobacterium was more abundant in tumors compared to normal tissue (Supplementary Figure S3A-B),

whereas a cognate Gram-negative oral commensal anaerobe, *Prevotella*, was not (Supplementary Figure S3C-D). Receiver operating characteristic (ROC) analysis identified a *Fusobacterium* RA cutoff of 0.016 (specificity: 92.7%; sensitivity: 28.8%). Patients with RA above the threshold had better OS and DSS (Supplementary Figure S4).

Next, we questioned whether any particular *Fusobac*terium species were associated with survival. Patients were stratified into groups with detectable and undetectable species (Supplementary Figure S5). In Cox regression, only *Fusobacterium nucleatum* detectability was significantly associated with OS (HR: 0.43 [95% CI: 0.19-0.97], p =0.042; Supplementary Figure S6). Kaplan-Meier modeling showed that *F. nucleatum* detectability was associated with improved OS (P < 0.001, Supplementary Figure S7A), with a trend for improved DSS (P = 0.096, Supplementary Figure S7B).

In multivariate Cox modeling with established predictors of survival (disease stage, smoking and Human Papilloma Virus [HPV] status), both *Fusobacterium* and *F. nucleatum* detectability were strongly associated with OS (P < 0.001 for both, Supplementary Figures S8A/S9A) and DSS (P < 0.001 and P = 0.015 for each respectively, Supplementary Figures S8B/S9B).

To test the validity of these results, we evaluated whether the abundance of *Fusobacterium* was also predictive of treatment efficacy in the separate MicroLearner cohort (n = 175; described in Supplementary Text and Supplementary Table S2) by dividing it into patient groups with *Fusobacterrium* RA either below (FusoLO) or above (FusoHI) the cohort median, as the commensal nature of *Fusobacterium* in the oral cavity makes detectability nearly universal in saliva [3]. We used progression-free survival (PFS) as an endpoint because, with a median followup of 33.6 months (range 4-57 months), very few deaths (n = 6, 3.4%) had occurred. FusoHI patients had a trend for better PFS (P = 0.054; Figure 1C). Only 10 events (15.6%) of progression were observed in patients with HPV-positive

Abbreviations: HNSCC, Head and Neck Squamous Cell Carcinoma; RA, relative abundance; OS, Overall Survival; CI, Confidence Interval; DSS, Disease-Specific Survival; HR, Hazard Ratio; TCMA, The Cancer Microbiome Atlas; TCGA, The Cancer Genome Atlas; PFS, Progression-Free Survival; HPV, Human Papillomavirus; OSCC, Oral Squamous Cell Carcinoma; MOI, Multiplicity of Infection; LDH, Lactate Dehydrogenase; ROC, Receiver Operating Characteristic; Fnuc, *Fusobacterium nucleatum*; Fper, *Fusobacterium periodonticum*; FnucHI, Group of patients where RA of Fnuc is above the cohort median; FnucLO, Group of patients where RA of Fper is above the cohort median; FperHI, Group of patients where RA of Fper is below the cohort median; FperLO, Group of patients where RA of Fper is below the cohort median; ATP, Adenosine triphosphate; 16S rRNA, 16S ribosomal RNA.

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oropharyngeal cancer, so we conducted a separate analysis including all patients except these ("HPVneg cohort"; n = 111, 29.7% event rate), where FusoHI patients had significantly better PFS (P = 0.011, Figure 1D). ROC analysis identified a salivary *Fusobacterium* RA cutoff of 2.760 in this cohort (specificity: 65.2%; sensitivity: 55.8%). Patients with RA above this threshold had better PFS (Supplementary Figure S10). *F. nucleatum* (Fnuc) and *F. periodonticum* (Fper) were the most abundant fusobacterial species. There was a non-significant trend for better PFS in FnucHI (*F. nucleatum* RA > median *F. nucleatum* RA) patients, but FperHI patients had significantly better PFS (P = 0.021; Supplementary Figure S11).

Given our clinical observations, we reasoned that Fusobacterium may contribute to HNSCC killing. We initially explored the effect of F. nucleatum on oral SCC (OSCC) evaluated with an ATP-based viability assay. TR146 cells were infected with F. nucleatum at multiplicity of infection (MOI) ranging from 0.5 to 5. With increasing MOI, a more significant reduction in OSCC cell viability was observed (Supplementary Figure S12A). Separate experiments using lactate dehydrogenase (LDH) activity and crystal violet assays validated these findings (Supplementary Figure S12B-C). We also tested whether the F. nucleatum medium caused any OSCC death if added without any previous contact with bacteria and confirmed that it did not (Supplementary Figure S12D). A significant decrease in viability was observed from 24h post-infection (Supplementary Figure S13).

To test whether the observed effects of *F. nucleatum* on OSCC cytotoxicity were strain-specific, cell-line specific and not a general characteristic of oral commensal anaerobes, we co-cultured multiple cell lines of OSCC (TR146, HN5 and HSC-3) with either of two *F. nucleatum* strains or *Prevotella oralis* (MOI = 100) and evaluated their effect on OSCC viability (Figure 1E), validated with a crystal violet assay (Supplementary Figure S14). *P. oralis*, like *F. nucleatum*, is an oral commensal Gram-negative anaerobe. *P. oralis* infection did not impact OSCC viability, while both *F. nucleatum* strains caused a reduction in OSCC viability. We next questioned whether other *Fusobacterium* species caused OSCC killing. We tested the effect of *F.*

periodonticum on OSCC cultures at MOI 100 and found that it caused OSCC killing similarly to *F. nucleatum* (Figure 1F). At lower MOI (0.5-5), OSCC killing was also overall similar between the two species and rose with MOI (Supplementary Figure S15). These results suggest that other *Fusobacterium* species which are phylogenetically close to *F. nucleatum*, but not all oral commensal Gram-negative anaerobes can cause OSCC killing.

We next asked whether OSCC killing was mediated by a surface protein or by secreted compounds/metabolites (Figure 1G). Firstly, OSCC cells were infected with F. nucleatum, which was either alive or heat-inactivated (inFnuc), and OSCC viability was assessed. We also tested whether the supernatant of F. nucleatum culture was sufficient to cause OSCC death. F. nucleatum supernatant caused OSCC killing, whereas fresh medium did not. Coculture of F. nucleatum washed in fresh broth significantly attenuated OSCC killing compared to growth broth, suggesting continued production of supernatant in co-culture. inFnuc caused OSCC killing only when added to co-culture with growth broth but not with fresh broth. Separately, we used transwell inserts to prevent direct contact of F. nucleatum with OSCC while allowing for any secreted molecules to move freely between them (Supplementary Figure S16). Significant cell killing was observed in transwell replicates, more substantially when F. nucleatum was in direct contact with OSCC, which may be attributable to higher local concentrations in direct contact co-culture compared to transwell replicates. Taken together, these results indicate that F. nucleatum mediates OSCC killing primarily via the bacterial secretome.

Although colorectal cancer studies indicate that *F. nucleatum* contributes to oncoprogression and treatment resistance, these bacteria are not common constituents of the normal intestinal microbiota, whereas they are common components of the normal oral microbiota [4]. Previous studies often assume that a higher tumoral abundance of *Fusobacterium*, which we also detected, indicates its oncogenic role [5]. However, our findings suggest that its presence may enhance HNSCC treatment efficacy. Limitations of this study are discussed in the Supplementary Text.

FIGURE 1 *Fusobacterium* detectability enhances survival in HNSCC, while its presence in co-culture is toxic for HNSCC. In the TCMA cohort, intra-tumoral *Fusobacterium* detectability determines better OS (A) and DSS (B). In the MicroLearner cohort, a salivary *Fusobacterium* RA above the cohort median determines better PFS in the full cohort (C) and in the HPV-neg cohort (D). (E) In 2D co-culture, OSCC toxicity with *F. nucleatum* (MOI 1) and lack thereof with *P. oralis* (NCTC 11459, MOI 1) is reproducible in the TR146, HN5 and HSC3 cell lines. Additionally, it is reproducible with different *F. nucleatum* strains (ATCC 23726, ATCC 25586; MOI 1). (F) *F. nucleatum* and *F. periodonticum* cause OSCC killing at similar magnitude. (G) OSCC viability in single culture with or without the supernatant of *Fnuc* culture, or fresh broth (i.e., broth without *Fnuc* supernatant), or *F. nucleatum* (either alive (Fnuc) or heat-inactivated (inFnuc)), showing that *Fnuc* supernatant and/or live *Fnuc* (i.e., producing supernatant molecules) are sufficient and necessary for OSCC toxicity. Significance levels: ns: not significant; *: P < 0.01; ***: P < 0.001; ****: P < 0.0001; ns: ###. Abbreviations: DSS, disease-specific survival; Fnuc, *F. nucleatum*; Fper, *F. periodonticum*; HNSCC, head and neck squamous cell carcinoma; inFnuc, heat-inactivated *F. nucleatum*; MOI, multiplicity of infection; OS, overall survival; OSCC, oral squamous cell carcinoma; PFS, progression-free survival.

In summary, our preliminary research suggests that *Fusobacterium* actively determines survival outcomes in HNSCC. Ongoing research will validate its role as a predictive biomarker in HNSCC and dissect the mechanism by which fusobacteria cause HNSCC killing.

AUTHOR CONTRIBUTIONS

Miguel Reis Ferreira conceived and designed the study. Miguel Reis Ferreira, Anjali Chander, Aize Pellon and David Moyes designed the experiments. Jacopo Iacovacci and Tiziana Rancati designed and analyzed MicroLearner study data. Miguel Reis Ferreira, Anjali Chander and Jacopo Iacovacci analyzed the data. Jacopo Iacovacci, Rhadika Kataria, Anita Grigoriadis, David Moyes and Tiziana Rancati supported data analysis. Anjali Chander, Jacopo Iacovacci, Tiziana Rancati and Miguel Reis Ferreira reviewed the results, interpreted the data and wrote the manuscript. Anjali Chander, Jacopo Iacovacci, Aize Pellon, Rhadika Kataria, Anita Grigoriadis, John Maher, Cynthia Sears, Gilad Bachrach, Teresa Guerrero Urbano, Mary Lei, Imran Petkar, Anthony Kong, Tony Ng, Ester Orlandi, Nicola Alessandro Iacovelli, Loris De Cecco, Mara Serena Serafini, David Moyes, Tiziana Rancati and Miguel Reis Ferreira critically reviewed the manuscript for important intellectual content and approved the final version. Miguel Reis Ferreira has primary responsibility for the final content of the manuscript. All authors reviewed and approved the final manuscript for submission.

ACKNOWLEDGMENTS

First, we thank the patients who donated samples for this research. We thank Prof William Wade (King's College London), Prof Martin Forster (University College London), Dr Olivier Martinez (King's College London) and Dr Joseph Aduse-Opoku (King's College London) for their insightful discussions and help with the methodology. We also thank Prof Mahvash Tavassoli (King's College London) who generously provided some of the cancer cell lines used in this research. We acknowledge funding from Wilson + Olegario: Philanthropy through Guys Cancer Charity, Guys Cancer Charity and Cancer Research UK through the City of London Cancer Centre. J.I. acknowledges funding from Fondazione Regionale per la Ricerca Biomedica, grant ID 2721017 (MicBioRadio).

CONFLICT OF INTEREST STATEMENT The authors declare no competing interests.

FUNDING INFORMATION

Wilson + Olegario: Philanthropy through Guys Cancer Charity (MRF) Guys Cancer Charity (MRF) Cancer Research UK through the City of London Cancer Centre (MRF) Fondazione Regionale per la Ricerca Biomedica, grant ID 2721017 (JI).

DATA AVAILABILITY STATEMENT

Data from the Cancer Microbiome and Cancer Genome Atlases are publicly available, as indicated in the text. All other data generated or analyzed during this study are included with the published article and its supplementary information files.

ETHICS APPROVAL AND CONSENT TO PARTICIPATE

The MicroLearner observational study of the microbiome in patients treated with radiotherapy for head and neck and prostate cancers was registered on ClinicalTrials.gov (ID: NCT03294122) and approved by the local Ethical Committee (ID INT 11/17). All patients provided written informed consent and agreed that incidental findings would not be disclosed to them or any clinician.

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SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.